

FREE SULPHITE (FSO₂)

(Manual, Auto-Analyser & Microplate Liquid Stable, Rapid, Format)

ASSAY PROCEDURE

K-FSULPH 08/17

(80 Manual Assays of each per Kit) or (800 Auto-Analyser Assays of each per Kit) or (800 Microplate Assays of each per Kit)



INTRODUCTION:

Sulphur dioxide (SO_2) is used widely as an additive in various forms, most commonly as sulphites (or sulfites), in the wine, beverage and food industries where it acts primarily as an antimicrobial and antioxidant preservative.

During wine making sulphites are used as an essential additive, usually at post-malolactic fermentation, in the control of contamination by Brettanomyces during aging and also to protect the wine against detrimental "oxidative and enzymatic browning". SO_2 is only active as an antimicrobial and antioxidant preservative in the unbound "free" form. Given that SO_2 becomes "inactive" when it binds the colour pigments of wine, and with legal restrictions on SO_2 levels in wine, it has become valuable to wine producers to measure both the Free SO_2 (FSO₂) and Total SO_2 (TSO₂). In addition to this, due to the increased awareness of the adverse effects of sulphites and the prevalence of sulphite intolerance in some individuals, sulphite levels in foods and drinks are strictly regulated by various governing bodies and therefore there is a requirement for accurate determination of the level of sulphites in foods, beverages and wines.

This kit (K-FSULPH) is suitable for the specific measurement of both "Free Sulphite", especially in wines, beverages, foodstuffs, and other materials.

PRINCIPLE:

The Free Sulphite assay is based on the reaction principle of SO_2 , fuchsin and aldehyde binding. As the reaction between FSO_2 and the aldehyde proceeds, the equilibrium reaction between "bound" SO_2 and FSO_2 shifts towards FSO_2 . With red wine this equilibrium reaction is linear.

SPECIFICITY, SENSITIVITY, LINEARITY AND PRECISION:

The smallest differentiating absorbance for the assay is 0.010 absorbance units. This corresponds to a FSO $_2$ concentration of ~ 1 mg/L of sample solution at a sample volume of 0.05 mL. The detection limit is ~ 2 mg/L, which is derived from an absorbance difference of 0.020 with a sample volume of 0.05 mL.

The assay is linear over the range of 0.25 to 7.5 µg of FSO $_2$ per assay. With a sample volume of 0.05 mL, this corresponds to a FSO $_2$ concentration of \sim 5 to 150 mg/L of sample solution. In duplicate determinations using one sample solution, an absorbance difference of 0.005 to 0.010 may occur. With a sample volume of 0.05 mL, this corresponds to a FSO $_2$ concentration of \sim 0.5 to \sim 1 mg/L of sample solution. If the sample is diluted during sample preparation, the result is multiplied by the dilution factor, F. If, in sample preparation, the sample is weighed, e.g. 10 g/L, a difference of 0.02 to 0.05 g/100 g can be expected.

INTERFERENCES:

Thiol reactive compounds (e.g. aldehydes and maleimide compounds) can interfere with FSO₂ assay.

Endogenous aldehydes in wine are not considered as an interference for the FSO₂ assay for wine samples.

SAFETY:

The general safety measures that apply to all chemical substances should be adhered to.

For more information regarding the safe usage and handling of this product please refer to the associated SDS that is available from the Megazyme website.

KITS:

Kits suitable for performing 80 assays in manual format (or 800 assays in auto-analyser format or 800 of each test in microplate format) are available from Megazyme. The kits contain the full assay method plus:

Bottle I: (x2) Free Sulphite Reagent I (40 mL)
Stable for > 18 months at room temperature.

Bottle 2: (x2) Free Sulphite Reagent 2 (20 mL)
Stable for > 18 months at room temperature.

Sodium Sulphite (5 g).

Stable for > 5 years at room temperature.

PREPARATION OF REAGENT SOLUTIONS (SUPPLIED):

I & 2. Use the contents of bottles I & 2 as supplied.

3. Use the contents of bottle 3 as described for the preparation of the FSO₂ standard and calibration curve.

RECOMMENDED REAGENTS (NOT SUPPLIED):

- I. Citric acid (Sigma cat. no. 251275).
- 2. Sodium Sulphite (Sigma cat. no. 71989).

EQUIPMENT (RECOMMENDED):

- I. Disposable plastic cuvettes (I cm light path, 3 mL).
- 2. Micro-pipettors, e.g. Gilson Pipetman® (200 µL and I mL).
- Positive displacement pipettor, e.g. Eppendorf Multipette[®]

 with 25 mL Combitip[®] [to dispense 0.5 mL and 1.0 mL aliquots of reagent solutions (solutions 1-4)].
- 4. Stop clock.
- 5. Analytical balance.
- 6. Spectrophotometer set at 575 nm.
- 7. Vortex mixer (e.g. IKA® Yellowline Test Tube Shaker TTS2).
- 8. Whatman No. I (9 cm) filter papers.

A. MANUAL ASSAY PROCEDURE

NOTES:

- For each batch of samples that is applied to the determination of FSO₂ either a single point standard or a calibration curve must be performed concurrently using the same batch of reagents.
- 2. Prepare the FSO₂ standards for the single point standard or full calibration curve as described for the Manual Assay Procedure.

Wavelength: 575 nm

Cuvette: I cm light path (glass or plastic)

Temperature: ~ 25°C **Final volume:** 2.55 mL

Sample solution: $0.25-7.50 \mu g$ of SO_2 per cuvette (5-150 mg/L)

(in 0.05 mL sample volume)

Read against air (without cuvette in the light path) or against water

Pipette into cuvettes	Sample	Sample Standard				
distilled water (at ~ 25°C) Free Sulphite Reagent I sample standard	1.00 mL 1.00 mL 0.05 mL	1.00 mL 1.00 mL				
Mix* and read absorbances of the solutions (A _I) after exactly 3 min. Then add:						
Free Sulphite Reagent 2 0.50 mL 0.50 mL 0.50 mL						
Mix* and read absorbances of the solutions (A2) after exactly 3 min.						
Mix* and read absorbances of the solutions (A3) after exactly 3 min.						

^{*} for example with a plastic spatula or by gentle inversion after sealing the cuvette with a cuvette cap or Parafilm®.

PREPARATION OF THE FSO₂ SINGLE POINT STANDARD:

The FSO_2 standard is required for the calculation of FSO_2 in the test samples.

Weigh I g of citric acid into a I L volumetric flask, make to I L with distilled water and dissolve. Accurately add 196 mg of sodium sulphite and dissolve. This is the 100 mg/L standard, use this directly as the single point standard in the Manual Assay Procedure for Free Sulphite (FSO₂). Prepare on the day of use. Stable for I day at room temperature.

NOTE: Alternatively, a full FSO_2 calibration curve can be used and the FSO_2 content calculated as described for "Preparation of the FSO_2 Calibration Curve".

CALCULATION OF FSO₂ CONTENT USING A SINGLE POINT STANDARD:

Determine the ΔA_{FreeSO_2} for the sample, standard and blank. Subtract the ΔA_{FreeSO_2} of the blank from that of the sample and the standard, thereby obtaining $\Delta A_{FreeSO_2\text{-SAMPLE}}$ and $\Delta A_{FreeSO_2\text{-STD}}$, respectively.

The value of $\Delta A_{FreeSO_2-SAMPLE}$ and ΔA_{FreeSO_2-STD} should be at least 0.100 absorbance units to achieve sufficiently accurate results.

The concentration of FSO₂ can be calculated as follows:

$$\Delta A_{FreeSO_2} = (A_2 - A_1) - (A_3 - A_2)$$

$$c = \frac{\Delta A_{FreeSO_2-SAMPLE}}{\Delta A_{FreeSO_2-STD} / c_{FreeSO_2-STD}} \times F$$
 [mg/L]

where:

c_{FreeSO₂-STD} = concentration of FSO₂ standard (mg/L) F = dilution factor

If the sample is diluted during preparation, the result must be multiplied by the dilution factor, F.

When analysing solid and semi-solid samples which are weighed out for sample preparation, the content (g/100 g) is calculated from the amount weighed as follows:

Content of FSO₂

 $= \frac{c_{FreeSO_2} [g/L \text{ sample solution}]}{\text{weight}_{sample} [g/L \text{ sample solution}]} \times 100 \quad [g/100 \text{ g}]$

NOTE: These calculations can be simplified by using the Megazyme *Mega-Calc*[™], downloadable from where the product appears on the Megazyme website (www.megazyme.com).

PREPARATION OF THE FSO₂ CALIBRATION CURVE:

Weigh I g of citric acid into a I L volumetric flask, make to I L with distilled water and dissolve. Accurately add 295 mg of sodium sulphite and dissolve. **This is the I50 mg/L FSO₂ solution.**

Prepare on the day of use. Stable for I day at room temperature.

Prepare the FSO_2 standard solutions as described in the table below and use directly as standards to determine A_2 in the Manual Assay Procedure for Free Sulphite (FSO_2).

Pipette into 13 mL polypropylene tubes	STD 0	STD I	STD 2	STD 3	STD 4	STD 5
(FSO ₂) mg/L	0	15	30	60	120	150
0.1% (w/v) citric acid (mL)	5.00	4.50	4.00	3.00	1.00	-
FSO ₂ solution (mL)	-	0.50	1.00	2.00	4.00	5.00
Total Volume (mL)	5.00	5.00	5.00	5.00	5.00	5.00

CALCULATION OF FSO₂ CONTENT USING THE FSO₂ CALIBRATION CURVE:

I. FSO₂ CALIBRATION CURVE ANALYSIS:

- I. Determine the absorbance (A₂) of each FSO₂ standard (STD 0-5). Subtract the absorbance of STD 0 from the absorbance of the other standards (STD 1-5), thereby obtaining ΔA_{FreeSO_2} (An example is given in Table 1).
- 2. Calculate M as follows, for each standard (STD 1-5):

$$M = \frac{FSO_2 (mg/L)}{\Delta A_{FreeSO_2}} [(mg/L)/\Delta A_{FreeSO_2}]$$

3. Calculate the mean M as follows:

mean M =
$$\frac{(M_{STD1} + M_{STD2} + M_{STD3} + M_{STD4} + M_{STD5})}{5} [(mg/L)/\Delta A_{FreeSO_2}]$$

Use "mean M" to calculate the FSO₂ content of the test samples in section 2 "Free Sulphite (FSO₂) Content".

Example:

Examples of the FSO_2 calibration curve calculations are given in Table I along with a graphical representation of a typical FSO_2 calibration curve (Figure 2).

FSO ₂ standard	FSO ₂ (mg/L)	A ₅₇₅	∆A _{Free} SO ₂	M [(mg/L)/ ∆A _{FSO2}]
STD 0	0	0.132	0.000	-
STD I	25	0.383	0.251	99.761
STD 2	50	0.628	0.496	100.806
STD 3	100	1.104	0.972	102.881
STD 4	125	1.364	1.232	101.469
STD 5	150	1.639	1.507	99.594
mean M	-	-	-	100.893

Table 1. Calculations for a typical FSO₂ calibration curve.

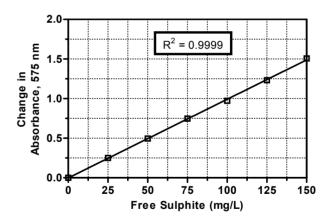


Figure 2. Calibration curve demonstrating the linearity of the colourimetric FSO₂ determination by K-FSULPH. The sulphite standards were analysed in a cuvette with a 1.0 cm light-path.

NOTE: These calculations can be simplified by using the Megazyme *Mega-Calc*[™], downloadable from where the product appears on the Megazyme website (www.megazyme.com).

2. FREE SULPHITE (FSO₂) CONTENT:

Determine the ΔA_{FreeSO_2} for the sample and blank. Subtract the ΔA_{FreeSO_2} of the blank from that of the sample, thereby obtaining ΔA_{FreeSO_2} -SAMPLE·

The value of $\Delta A_{FreeSO_2-SAMPLE}$ should be at least 0.100 absorbance units to achieve sufficiently accurate results.

As the reaction between FSO_2 and the aldehyde proceeds, the equilibrium reaction between "bound" SO_2 and FSO_2 shifts towards FSO_2 . With red wine this equilibrium reaction is linear and the concentration of FSO_2 can be calculated as follows:

$$\Delta A_{FreeSO_2}$$
 = $(A_2 - A_1) - (A_3 - A_2)$
c = mean M x F x $\Delta A_{FreeSO_2-SAMPLE}$ [mg/L]

where:

mean M = mean value of FSO_2 standards $[(mg/L)/\triangle A_{FreeSO_2}]$ = dilution factor

If the sample is diluted during preparation, the result must be multiplied by the dilution factor, F.

When analysing solid and semi-solid samples which are weighed out for sample preparation, the content (g/100 g) is calculated from the amount weighed as follows:

Content of FSO₂

$$= \frac{c_{FreeSO_2} [g/L \text{ sample solution}]}{\text{weight}_{sample} [g/L \text{ sample solution}]} \times 100 \quad [g/100 \text{ g}]$$

NOTE: These calculations, including the calibration curve analysis, can be simplified by using the Megazyme *Mega-Calc*[™], downloadable from where the product appears on the Megazyme website (www.megazyme.com).

B. AUTO-ANALYSER ASSAY PROCEDURE FOR FREE SULPHITE (FSO₂):

NOTES:

- I. The Auto-Analyser Assay Procedure for FSO₂ can be performed using either a single point standard or a full calibration curve.
- For each batch of samples that is applied to the determination of FSO₂ either a single point standard or a calibration curve must be performed concurrently using the same batch of reagents.
- 3. Prepare the FSO₂ standards for the single point standard or full calibration curve as described for the Manual Assay Procedures for FSO₂, respectively.
- 4. To measure FSO₂ the auto-analyser must have a slope correction capability.

Wavelength: 575 nm Calculation: Kinetic

Temperature: ~ 25°C or 37°C **Reaction:** Absorbance increase

Final volume: 0.255 mL

Linearity: $0.025-0.750 \mu g \text{ of SO}_2 \text{ per cuvette } (5-150 \text{ mg/L})$

(in 0.005 mL sample volume)

Pipette into cuvettes	Sample	Standard			
Free Sulphite Reagent I distilled water sample standard	0.100 mL				
Read absorbances of the solutions (A ₁) after exactly 3 min. Then add:					
Free Sulphite Reagent 2 0.050 mL 0.050 mL					
Read absorbances of the solutions (A ₂) after exactly 3 min.					
Read absorbances of the solutions (A ₃) after exactly 3 min.					

CALCULATION FORMULA FOR FSO₂ (AUTO-ANALYSER):

$$(A_2 - A_1) - (A_3 - A_2)$$

C. MICROPLATE ASSAY PROCEDURE:

NOTES:

- I. The Microplate Assay Procedure for FSO₂ can be performed using either a single point standard or a full calibration curve.
- For each batch of samples that is applied to the determination of FSO₂ either a single point standard or a calibration curve must be performed concurrently using the same batch of reagents.
- Prepare the FSO₂ standards for the single point standard or full calibration curve as described for the Manual Assay Procedures for FSO₂, respectively.
- 4. Calculation of FSO₂ content should be performed as described for the Manual Assay Procedures.
- The calculation of FSO₂ content, including the calibration curve analysis, can be simplified by using the Megazyme Mega-Calc™, downloadable from where the product appears on the Megazyme website (www.megazyme.com).

Wavelength: 575 nm

Microplate: 96-well (e.g. clear flat-bottomed, glass or plastic)

Temperature: ~ 25°C or 37°C

Final volume: 0.255 mL

Linearity: $0.025-0.750 \mu g \text{ of SO}_2 \text{ per cuvette } (5-150 \text{ mg/L})$

(in 0.005 mL sample volume)

Read against air (without cuvette in the light path) or against water

Pipette into cuvettes	Sample	Standard	Blank		
distilled water (at ~ 25°C) Free Sulphite Reagent I sample standard	0.100 mL		0.105 mL 0.100 mL - -		
Mix* and read absorbances of the solutions (A _I) after exactly 3 min. Then add:					
Free Sulphite Reagent 2 0.050 mL 0.050 mL 0.050 mL					
Mix* and read absorbances of the solutions (A2) after exactly 3 min.					
Mix* and read absorbances of the solutions (A ₃) after exactly 3 min.					

^{*} for example using microplate shaker, shake function on a microplate reader or repeated aspiration (e.g. using pipettor set at 50-100 μ L volume).

EQUIPMENT FOR MICROPLATE ASSAY FORMAT (RECOMMENDED):

- Disposable 96 well polystyrene clear, flat bottom microplates, e.g. Matrix Technologies Corp. cat. no. 4915 (www.matrixtechcorp. com).
- 2. Disposable 25 mL reagent reservoirs, e.g. Matrix Technologies Corp. cat. no. 809311 (www.matrixtechcorp.com).
- 3. Micro-pipettors, e.g. Gilson Pipetman® (200 μ L and I mL) and Multichannel Micro-pipettors, e.g. Gilson Pipetman® Ultra 8-channel (I-20 μ L and 20-300 μ L).
- 4. Stop clock.
- 5. Analytical balance.
- 6. Microplate shaker, e.g. Heidolph Titramax 100 or 1000 (www. heidolph-instruments.com).
- 7. Microplate reader set at 575 nm.
- 8. Vortex mixer (e.g. IKA® Yellowline Test Tube Shaker TTS2).
- 9. Whatman No. I (9 cm) filter papers.

SAMPLE PREPARATION:

I. Sample dilution.

The amount of FSO $_2$ present in the cuvette (i.e. in the 0.05 mL of sample being analysed) should range between 0.25 and 7.5 μg . The sample solution must therefore be diluted sufficiently to yield a concentration between 5 and 150 mg/L.

Dilution table (FSO₂)

Estimated concentration of FSO ₂ (mg/L)	Dilution with water	Dilution factor (F)
< 120 120-1200 > 1200	No dilution required + 9 + 99	1 10 100

If the value of ΔA_{FreeSO_2} is too low (e.g. < 0.100), weigh out more sample or dilute less strongly.

2. Sample clarification.

a. Solutions:

Carrez I solution. Dissolve 3.60 g of potassium hexacyanoferrate (II) $\{K_4[Fe(CN)_6].3H_2O\}$ (Sigma cat. no. P9387) in 100 mL of distilled water. Store at room temperature.

Carrez II solution. Dissolve 7.20 g of zinc sulphate (ZnSO $_4$.7H $_2$ O) (Sigma cat. no. Z4750) in 100 mL of distilled water. Store at room temperature.

Sodium hydroxide (NaOH, 100 mM). Dissolve 4 g of NaOH in I L of distilled water. Store at room temperature.

b. Procedure:

Pipette the liquid sample into a 100 mL volumetric flask which contains approx. 60 mL of distilled water, or weigh sufficient quantity of the sample into a 100 mL volumetric flask, and add 60 mL of distilled water. Carefully add 5 mL of Carrez I solution, 5 mL of Carrez II solution and 10 mL of NaOH solution (100 mM). Mix after each addition. Fill the volumetric flask to the mark, mix and filter.

3. General considerations.

- (a) Liquid samples: clear, slightly coloured and approximately neutral, liquid samples can be used directly in the assay.
- **(b)** Acidic samples: if > 0.1 mL of an acidic sample is to be used undiluted (such as red wine or coloured fruit juice), the pH of the solution should be increased to approx. 7.4 using 2 M NaOH, and the solution incubated at room temperature for 30 min.

- (c) Carbon dioxide: samples containing carbon dioxide should be degassed by increasing the pH to approx. 7.4 with 2 M NaOH and gentle stirring, or by stirring with a glass rod.
- (d) Strongly coloured samples: if used undiluted, strongly coloured samples should be treated by the addition of 0.2 g of polyvinylpolypyrrolidone (PVPP)/10 mL of sample. Shake the tube vigorously for 5 min and then filter through Whatman No. I filter paper.
- (e) **Solid samples:** homogenise or crush solid samples in distilled water and filter if necessary.
- (f) Samples containing fat: extract such samples with hot water at a temperature above the melting point of the fat, e.g. in a 100 mL volumetric flask at 60°C. Adjust to room temperature and fill the volumetric flask to the mark with distilled water. Store on ice or in a refrigerator for 15-30 min and then filter. Discard the first few mL of filtrate and use the clear supernatant (which may be slightly opalescent) for assay.
- (g) Samples containing protein: deproteinise samples containing protein with Carrez reagents, alternatively use ice cold perchloric acid. Add an equal volume of ice-cold I M perchloric acid with mixing. Filter or centrifuge at 1,500 g for 10 min and adjust the pH of the supernatant to between 7 and 8 with I M KOH. Use the supernatant in the assay after appropriate dilution.

SAMPLE PREPARATION EXAMPLES:

(a) Determination of FSO₂ in white wine.

The FSO_2 concentrations of white wine can generally be determined without any sample treatment (except dilution according to the dilution table). Typically, no dilution is required and a sample volume of 0.05 mL is satisfactory.

(b) Determination of FSO₂ in red wine.

The FSO_2 concentrations of red wine can generally be determined without any sample treatment (except dilution according to the dilution table). Typically, a dilution of 1:2 and sample volume of 0.05 mL are satisfactory.

(c) Determination of FSO₂ in beer.

After removal of carbon dioxide by stirring with a glass rod, dilute the sample according to the dilution table and analyse. Typically, no dilution is required and a sample volume of 0.05 mL is satisfactory.

(d) Determination of FSO₂ in fruit juice.

Dilute the sample to yield a FSO_2 concentration of less than 150 mg/L (see dilution table). Clear, neutral solutions can generally be determined without any sample treatment (except dilution). Turbid liquids generally only require centrifuging or filtering before the

dilution step. Coloured solutions are usually suitable for analysis after dilution to an appropriate FSO_2 concentration. However, if coloured solutions require analysis undiluted, they may need decolourising as follows: adjust 25 mL of liquid sample to approx. pH 7.4 with 1 M NaOH and increase the volume to 50 mL with distilled water. Add 0.5 g of PVPP, stir for 5 min and filter through Whatman No. I filter paper or centrifuge. Use the clear, slightly coloured filtrate or supernatant directly in the assay. Typically, no dilution is required and a sample volume of 0.05 mL is satisfactory.

(e) Determination of FSO₂ in potatoes.

NOTES:

Mince approx. 50 g of potato with 50 mL of 1 M potassium phosphate buffer (pH 4) and 0.1 mL of *n*-octanol (to minimise foaming) with a household mixer for approx. I min. Adjust to approximately pH 4 with 2 M KOH. Quantitatively transfer the mixture to a 500 mL volumetric flask with distilled water, fill up to mark with distilled water, mix and filter through Whatman No. I filter paper or centrifuge. *Typically, no dilution is required and a sample volume of 0.05 mL is satisfactory.*

NOTES:		



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